

DOI: 10.14744/ejmo.2022.51601 EJMO 2022;6(1):1–11

Review



Immunotherapy for Cancer: Strategies of Immunomodulation Therapy in Combination with Conventional Approaches

^(b) Tanvi Gupta,¹ ^(b) Mohd Murtaza,² ^(b) Mrinmoy Ghosh,³ ^(b) Wen Pin Su,^{1,4} ^(b) Narendra Kumar Pandey,⁵ ^(b) Umesh Goutam⁶

¹Institute of Clinical Medicine, College of Medicine, National Cheng Kung University, Tainan, Taiwan
²Microbial Biotechnology Division, CSIR- Indian Institute of Integrative Medicine, Jammu, Jammu & Kashmir, India
³Kalinga Institute of Industrial Technology, Bhuvneshwar, India
⁴Departments of Oncology and Internal Medicine, National Cheng Kung University Hospital, College of Medicine, National Cheng Kung University, Tainan, Taiwan
⁵School of Pharmaceutical Sciences, Lovely Professional University, Punjab, India
⁶Department of Molecular Biology and Genetic Engineering, School of Biosciences and Bioengineering, Lovely Professional University, Punjab, India

Abstract

The disease that poses a major threat to human life is cancer. Although different treatment techniques such as chemotherapy, radiotherapy, and chemically driven drugs are used, they do not show expected results and cause many side effects, eventually leading to the death of patients. However, there is one approach that is promising is the consolidation of cancer vaccines and immunotherapy, in which tumor-specific antigens, tumor-associated antigens, antigen-presenting cells, and toll-like receptors play a major role. The approach involves vaccines that are approved by the FDA and has shown good results in the latest research studies.

Keywords: Antigen-presenting cells, cancer vaccines, immunotherapy

Cite This Article: Gupta T, Murtaza M, Ghosh M, Su W, Pandey N, Goutam U. Immunotherapy for Cancer: Strategies of Immunomodulation Therapy in Combination with Conventional Approaches. EJMO 2022;6(1):1–11.

Cancer, which spreads at a faster rate, poses a great threat to the people in both developing and nondeveloping countries. There are achievements as well as failures in the fight against the prevention of cancer across the world.^[1] The goal is to prevent it permanently. It could become potent when the assets are limited, and there is a huge divergence in the initial stage detection, screening, and anticancer therapies.^[2] In 2018, 18.1 million new cases and 9.6 million deaths due to cancer were reported across the world. The rates of occurrence and death vary between generations and countries. Overall, over 9.6 million human population dies of cancer every year across the world, and the rapid rate of increase in the new cases will end up in identifying 22 million new cases every year in another 20 years. Among the different types of cancer, the major prevailing cancer types are lung, breast cancer in females, prostate, and colorectal cancer. Of these, the most frequent and major cause of death is due to lung cancer followed by liver, stomach, breast, and colorectal cancer.^[3]

Deaths due to different types of cancer have been reported in the literature: prostate,^[4] breast,^[5] colorectal,^[6] lung,^[7] liver,^[8] thyroid,^[9] and pancreatic.^[10] Factors contributing to

Phone: +91-9041329877 E-mail: umesh.14691@lpu.co.in, umeshbiotech@gmail.com

Submitted Date: August 11, 2021 Accepted Date: January 27, 2022 Available Online Date: March 09, 2022

[®]Copyright 2022 by Eurasian Journal of Medicine and Oncology - Available online at www.ejmo.org

OPEN ACCESS This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License.



Address for correspondence: Umesh Goutam, MD. School of Biosciences and Bioengineering, Lovely Professional University, Jalandhar, Punjab, India, 144411

cancer are alterations in genes, inherited genetic defects, age, gender, environmental exposure (e.g., UV rays, chemicals/preservatives used in the food, and radioactive materials), lifestyle, and variations leading to mutations. Chemotherapy,^[11, 12] radiotherapy,^[13, 14] chemically driven drugs,^[15] surgery,^[16] and certain inhibitors like vemurafenib against BRAF mutant skin cancer^[17] are a few clinical treatment methods.

Many studies have reported that an immune system can trigger progressive tumors immediately irrespective of the virulent factors. In renal and skin cancer, tumor-infiltrating lymphocytes inside the tumor show positive prognostic response,^[18, 19] in colorectal cancer, CD8+ T cells show positive prognostic response, and in breast and ovarian cancer, tumor-infiltrating lymphocytes show positive prognostic response.^[20]

Vaccines for cancer have a set of procedures that need to be considered for creation, multiplication, and promotion of immunity against tumors. In the last 100 years, more vaccines have been therapeutically applied to treat cancer and to control tumor antigens, antigen-presenting cells (APCs), and other immune signals of the tumor. The combination of cancer vaccines with conventional therapeutic approaches may lead to removing regulatory adoptive T cell suppression and improved clinical efficacy through co-stimulatory pathways. In particular, the combination approach can lead to activation of the immune modulate cells by rebooting the immune system, thus rendering tumor cells would be more susceptible to immune-mediated killing.^[21]

Mechanism of Therapeutic Vaccines for Cancer

The main objective of the vaccines against cancer is the activation of CD8+ cytotoxic T cells as the ongoing research on mice upholds the therapy by these cells. The APCs capture the neoantigens from the vaccine and the dead cancer cells. Then, the activated APCs migrate toward the lymph nodes, and major histocompatibility complex (MHC) molecules exhibit the neoantigens to T-lymphocytes. The CD4+ T cells build up immunity contrary to cancers, and CD8+ cytotoxic T lymphocytes help in the direct killing of the cancer cells by degranulation process using granzyme and perforin.^[22] The peculiar type of vaccination approach is the activation of CD8+ cytotoxic T cells linked with MHC class I. Vaccines are categorized in Figure 1 according to their modes of action.

Peptide Vaccines in Anticancer Therapy

A familiar way of vaccination against cancer is the transmission of the MHC class I antigenic determinants from tumorassociated antigens (TAA) to stimulate CD8+ T cell duplicates that can work against self-antigens.^[23] The peptides



Figure 1. Different types of vaccines and their mode of action.

that are added inside the adjuvants, such as Montanide, which is similar to incomplete Freund's adjuvant (IFA), in the presence or absence of cytokines including granulo-cyte-macrophage colony-stimulating factor (GM-CSF), interferon γ , and toll-like receptors (TLR) have shown feedback partially or completely in different phases of clinical trials.^[24]

Vaccines with one or more peptides can be infused with an adjuvant such as Montanide ISA-51 that is associated with cytokines such as GM-CSF to trigger APCs. This adjuvant has triggered TAA, in particular cytotoxic T cells. Cancer vaccines used in clinical trials are listed in Table 1. There is one challenge as IFA leads to the aggregation of T cells at the location of vaccination instead of promoting systemic immune response. Peptide vaccines are generally approved. The adjuvants and assembly of cancer vaccines are still ongoing. The advantages of peptide vaccines are they are easy to access, economically available at the mass level, and easy to be transported due to their stability.^[25]

The second peptide that can show potent clinical effectiveness is synthetic peptide, which consists of MHC class I and MHC class II antigenic determinants.^[26] A long peptide chain of length 23–45 amino acids infused subcutaneously proved efficient due to its processing and delivery pathway which triggers T cells.^[27]

Vaccines from Antigen-Presenting Cells in Oncoimmunity Therapy

Studies on different sets of APCs such as activated B cells and peripheral blood mononuclear cells have shown great signs of progress and advancement. Dendritic cells have a mixed population of APCs, adopting antigens to suit their environment. Further, they prepare and display these antigens to CD4+ and CD8+ T cells and integrate immune response signals to counteract the secretion of cytokines, such as interleukin 12 (IL-12) which alter them to type 1 immune re-

Table 1. Cancer vaccines (2)	015 onward) used in clinical t	trials				
Vaccine name	Targeted site of cancer	Functional malignant	Commercial producer	Current status	References	Web links
NeuVax	Breast cancer	Breast cancer with low to intermediate HER2 expression	Galena Biopharma, Inc.	Phase III	NCT01479 244	https://clinicaltrials.gov/ct2/show/NCT01479244
AVX 901	HER2+ cancer	Advanced or metastatic malignancies that express HER2	H. Kim Lyerly, Duke University	Phase I complete	NCT01526 473	https://clinicaltrials.gov/ct2/show/NCT01526473
INO 1400	Breast cancer	Multiple	Inovio Pharmaceuticals	Phase I complet e	NCT02960 594	https://clinicaltrials.gov/ct2/show/NCT02960594
GVAX	Pancreatic cancer	Metastatic pancreatic adenocarcinoma	Aduro Biotech, Inc.	Clinical phase II	NCT02004 262	https://clinicaltrials.gov/ct2/show/NCT02004262
SV-BR-1-GM	Metastatic breast cancer	Organ specific	BriaCell	Clinical phase II	NCT04418 219	https://www.clinicaltrials.gov/ct2/show/NCT04418219
DNA vaccine	Pancreatic cancer	Organ specific	Washington University School of Medicine	Clinical phase I	NCT03122 106	https://clinicaltrials.gov/ct2/show/NCT03122106
INO-5401 and INO-9012	Glioblastoma tumor	Glioblastoma (GBM)	Inovio Pharmaceuticals	Clinical phase 1/ll	NCT03491 683	https://clinicaltrials.gov/ct2/show/NCT03491683
IFx-Hu2.0	Cutaneous melanoma	Specific	Morphogenesis, Inc.	Early phase 1	NCT03655 756	https://clinicaltrials.gov/ct2/show/NCT03655756
PIVOT-RCC	Renal cell carcinoma	Specific	Washington University School of Medicine	Clinical phase II	NCT03598 816	https://clinicaltrials.gov/ct2/show/NCT03598816
INVAC-1	Advanced solid tumors	Hematologic malignancies and solid tumors	Invectys	Clinical phase l	NCT02301 754	https://clinicaltrials.gov/ct2/show/NCT02301754
VGX-3100	Anal neoplasm	Anal/perianal high grade squamous intraepithelia lesion (HSIL)	Inovio Pharmaceuticals	Clinical phase II	NCT03499 795	https://clinicaltrials.gov/ct2/show/NCT03499795
INO-5401 + INO-9012	Urothelial carcinoma	Metastatic/recurrent urothelial carcinoma (UCa)	Inovio Pharmaceuticals	Clinical phase I/II	NCT03502 785	https://clinicaltrials.gov/ct2/show/NCT03502785
pTVG-HP	Prostate cancer	Nonmetastatic, noncastrate prostate cancer	University of Wisconsin, Madison	Clinical phase II	NCT03600 350	https://clinicaltrials.gov/ct2/show/NCT03600350
PROST VAC-V/F	Metastatic hormone- sensitive prostate cancer	Prostate	Washington University School of Medicine	Clinical phase l	NCT03532 217	https://clinicaltrials.gov/ct2/show/NCT03532217
VB10.16 immunotherapy	High grade cervical intraepithelial neoplasia	High grade HPV16+ cervical intraepithelial	Vaccibody AS	Clinical phase I/II	NCT02529 930	https://clinicaltrials.gov/ct2/show/NCT02529930
GX-188E	Cervical intraepithelial neoplasia	neoplasia (HSIL; CIN2/3) Cervical intraepithelial neoplasia 2, 2/3, or 3 (CIN3)	Genexine, Inc.	Clinical phase II	NCT02596 243	https://clinicaltrials.gov/ct2/show/NCT02596243
MEDI0457	Head and neck cancer	Metastatic head and neck squamous cell cancer	Medlmmune LLC	Clinical phase l	NCT03162 224	https://clinicaltrials.gov/ct2/show/NCT03162224
VGX-3100	Cervical dysplasia	Multiple	Inovio Pharmaceuticals	Clinical phase III	NCT03185 013	https://clinicaltrials.gov/ct2/show/NCT03185013
GX-188E	Cervical cancer	Advanced, nonresectable HPV-positive cervical cancer	Genexine, Inc.	Clinical phase I/II	NCT03444 376	https://clinicaltrials.gov/ct2/show/NCT03444376
HPV DNA plasmid therape utic vaccine VGX-3100	Anal intraepithelial neoplasia	Human immunodeficiency virus (HIV)-positive	AIDS Malignancy Consortium high grade anal lesions	Clinical phase II	NCT03603 808	https://clinicaltrials.gov/ct2/show/NCT03603808
INO-3112 (MEDI0457)	Cervical cancer	Multiple	M.D. Anderson Cancer Center	Clinical phase II	NCT03439 085	https://clinicaltrials.gov/ct2/show/NCT03439085
VGX-3100, placebo	Cervical dysplasia	High grade squamous intraepithelial lesions (HSIL) of the cervix, associated with HPV-16 and/or HPV-18	Inovio Pharmaceuticals	Clinical phase III	NCT03721 978	https://clinicaltrials.gov/ct2/show/NCT03721978

_

name Targeted site of cancer Fu lized polyepitope Triple negative Edite Edite					
oolyepitope Triple negative breast cancer h-A Breast cancer A-A Breast cancer HER OX2/CDH3/MDM2 Breast cancer Nonrr 3P2-HER2-IGF1R Breast cancer Nonrr NA vaccine Triple negative breast cancer	unctional malignant	Commercial producer	Current status	References	Weblinks
In-A Breast cancer HER SOX2/CDH3/MDM2 Breast cancer HER BP2-HER2-IGF1R Breast cancer Nomm NA vaccine Triple negative breast cancer breast cancer	Organ specific	Washington University School of Medicine	Clinical phase l	NCT02348 320	https://clinicaltrials.gov/ct2/show/NCT02348320
SOX2/CDH3/MDM2 Breast cancer HER EBP2-HER2-IGF1R Breast cancer Nomm DNA vaccine Triple negative breast cancer	Organ specific	Washington University School of Medicine	Clinical phase l	NCT02204 098	https://clinicaltrials.gov/ct2/show/NCT02204098
FBP2-HER2-IGF1R Breast cancer Nonm DNA vaccine Triple negative breast cancer	R2-negative stage III–IV I breast cancer	Jniversity of Washington	Clinical phase l	NCT02157 051	https://clinicaltrials.gov/ct2/show/NCT02157051
DNA vaccine Triple negative breast cancer	metastatic, node positive, I HER2 negative	Jniversity of Washington	Clinical phase l	NCT02780 401	https://clinicaltrials.gov/ct2/show/NCT02780401
	Organ specific	Washington University School of Medicine	Clinical phase l	NCT03199 040	https://clinicaltrials.gov/ct2/show/NCT03199040
Prostate cancer	Organ specific	University of Wisconsin, Madison	Clinical phase l	NCT02411 786	https://clinicaltrials.gov/ct2/show/NCT02411786
Prostate cancer	Hormone-resistant prostate cancer	University of Wisconsin, Madison	Clinical phase l	NCT02499 835	https://clinicaltrials.gov/ct2/show/NCT02499835

sponse, tumor necrosis factor (TNF), interferon γ , and IL-2, and improve the stimulation of CD8+ cytotoxic T cells.^[28]

Vaccines from Dendritic Cells in Tumor Immunity

The clinical trials of these vaccines have proved to be uncommon as it involves ascertaining ways for vaccination. It is challenging to contrast clinical trials and analyze results regarding the efficiency of the trials. The tests have been performed on CD34+ progenitor cells, monocytes, tumorspecific antigens, TAA, and MHC class I peptides. These vaccines are infused inside the patient's body through the skin, blood, and lymph nodes. The advantages of these vaccines are they are cost-effective, nonhazardous, and they show good immune response. Suppression of tumors can also be seen in patients. The effective response of the clinical trials and immunology have been shown by dendritic cells harmonized with mucin 1-derived peptide and a mixture of PADRE peptides infused through the skin in patients suffering from renal cell carcinoma.^[29]

The clinical trials performed on patients with skin and thyroid cancer are MART-1,^[17] allogeneic tumor lysis,^[30] autologous tumor lysis,^[31] and transfection with RNA.^[32] Those performed on patients with kidney cancer and breast cancer are vibrations with peptides^[33] and fusion of allogeneic dendritic cells with autologous tumor.^[34] For multiple melanomas, the clinical trials used are vibrations with carcinoembryonic antigen (CEA) peptide^[35] and vibrations with mannan MUCI fusion protein.^[36] For modification in pox, virus encoding CEA with Tricom is used.^[37]

Modified Tumor-Based Vaccines

In previous studies, mice were vaccinated with destroyed tumor cells and transformed to show activation of immune cytokines such as GM-CSF.^[38] The major role was played by tumor-specific CTLs, which investigated the cDNA libraries formed from tumor cell-derived mRNA and transfection took place in the MHC molecule of the recipient. This can be achieved by focusing on the T cell antigens, where the screening of peptides from MHC molecules takes place by the use of mass spectrometry and reversed-phase high performance liquid chromatography. The development of vaccines established on autologous tumor cells is achievable but complicated.^[39]

Cell Line-Based Vaccines

Tests have been performed on allogeneic cell lines in the presence or absence of autologous tumor cells. The tumor cells explicitly increase GM-CSF, also known as G-Vax, which serves as an ultimate boost in the study where the patients having pancreatic cancer obtains recombinant listeria bacteria signifies the tumor associated antigen mesothelin in

presence or absence of G-Vax consisting of allogeneic pancreatic cancer cell lines.^[40] Numerous vaccines are desirable without any hindrance from induced antibody and incorporation of bacteria present to act as abundant features of natural infection by activation of TLR and foreign pathogen receptors.^[41]

Autologous Tumor Cell Vaccines for Immunotherapy

The cells can be taken into account for the transfection of APCs such as autologous or allogeneic cell lines with the genomic DNA of tumor. In this way, the undefined mutated genes in particular to tumor can be manufactured and conferred for triggering immune response. These vaccines are tedious to achieve from the patients who underwent surgery for a particular disease. The drawback is the production is limited to 2–3 doses of vaccines from the autologous tumor, and when there is availability of the autologous tumor, there is no consent about the processing, preservation, modification, and delivery for a candidate vaccine.^[42]

Virus-Mediated Vaccines in Oncolytic Immunotherapy

Vaccines such as Gardasil and Cervarix used against human papillomavirus are certified against the virus. Their performance takes place by triggering humoral immunity in contrast to viral capsid proteins inside noncontagious virallike particles. Adenoviruses can be treated as vectors precisely by infusing tumor antigens inside the muscle tissue. ^[43] These viruses are used in vivo to transform antigens into APCs and every virus shows rare results on the transformed cells from triggering to suppression of cells.^[44] A favorable approach that has been approved is GM-CSF which acts as an adjuvant or as APC transformed growth factor inside the herpes virus vectors. The commonly used vectors such as T-Vec have been recommended for patients against skin cancer in phase III trials.^[45] The clinical trials performed on the patients so far are: heterologous booster poxvirus tyrosinase for skin cancer^[46] and poxvirus encoded 5T4,^[47,42] heterologous booster poxvirus PSA and Tricom,^[48] and poxvirus-encoded CEA and Tricom for kidney and colorectal cancer.[49]

Other vaccines that have been used in clinical trials are as follows. For skin cancer: NY-ESO-1 and Iscomatrix,^[50] ganglioside, and IFA; for lung cancer: α GalCer PBMC with Interleukin-2 and GM-CSF,^[51] transduction of allogeneic tumor with antisense TGF- β 2,^[52] and transduction of allogeneic GM-CSF mixed with autologous tumor;^[53] for multiple melanomas: umbilical vein endothelial cells;^[54] for pleura cancer: autologous tumor with GM-CSF;^[55] for brain cancer: transduction of autologous tumor with antisense TGF- β 2;^[56] and for head and neck cancer: Hsp65.^[57]

Other techniques employed for the treatment of cancer are tumor ablation, where the removal of large and small tumors takes place; radiofrequency ablation, which involves heating at particular locations, leading to inflammation and necrosis, and triggers the activity of natural killer cells;^[58] and cryoablation, which involves the discharge of TAA, enhancing the immune response against tumors.^[59]

One example of therapeutic vaccine against cancer is sipuleucel-T produced by Dendreon, which has been certified by the Food and Drug Administration (FDA). The therapeutic mechanism of the cancer vaccine is shown in Figure 2. Therapeutic vaccines have been authorized for the analysis of metastatic prostate cancer in the long-term survival in phase III clinical trials.^[60]

How Does a Cancer Vaccine Work?

The definite responsive immunotherapy aims to trigger an immune response against the tumor by transmitting tumor antigens into dendritic cells and contributing the optimum requirements for the maturation of the dendritic cells inside an effective immune response of APCs. The four major steps describing the working of cancer vaccines are identification of tumor-rejection antigens, stimulation of a robust host's immune system, reducing the risk of autoimmunity, and evasion of the immune system (Fig. 3).

Identification of Tumor-Rejection Antigens

Tumor antigens are extracted from the cDNA library or from peptides as tumor-specific cytolytic T cells. The efficacy of the tumor antigens relies upon the prevalence and avidity of the T cells present inside the patient's body.



Figure 2. Therapeutic mechanism of vaccine for cancer treatment. **(a)** Various composition of antigen-specific cancer vaccine to deliver tumor antigen, **(b)** neoantigen taken by APCs at the vaccination site and then migrate to the lymph node, **(c)** antigen presentation by activated APCs to T cell through MHC-I and MHC-II and stimulation of B cell proliferation by T helper cell, **(d)** activation of antigen-specific CD4 helper and CD8 cytotoxic T cell leading to clonal expansion and migration to the tumor site, **(e)** killing of tumor cells.



Figure 3. Role of cancer vaccine response to the immune system inside the body.

^[61] Antigens vary in their efficacy in achieving immunotherapy. Very little or no tolerance is observed in tumor antigens related to fetal genes observed in immunological sites such as CEA, and MAGE family creates great tumor-rejection antigens. In the case of tissue-originated sites, MART1, ERBB2, and SILV show tolerance but uncertain tumor rejection antigens.^[62] The major types of tumor antigens strengthen with the description of telomerase reverse transcriptase, which acts as a potent antigen in patients suffering from cancer. Few other tumor antigens are survivin and OFA. These antigens prove to be key for safeguarding oncogenic traits of the tumor cells, where immune dodging can be hindered.^[63]

Stimulation of a Robust Host's Immune System

The objectives are to direct tumor antigens inside the dendritic cells and make the dendritic cells process antigens into robust stimulation of immune response. There are two pathways for dendritic cells: in vivo and ex vivo. The in vivo technique involves infusion of antigen combined with adjuvant inside the patient's body. It is easily understandable and most favorable. In the case of in vivo technique, dendritic cells are manipulated, and the loading of antigen will show the best distinction of action of the APCs. The research study shows that dendritic cell immunotherapy is efficient compared with other techniques. The CD4+ T cells provide immunity against tumor, cytokines such as interferon y helps in the stimulation of tumor cells toward CTL lysis, enhancing MHC class I interpretation and internal pathway, which trigger the innate arm of an antibody at the location of tumor and hinder angiogenesis.^[64] The CD8+ T cells consist of the effectors' arm of an antibody against the tumor reaction, from the information to reach an optimum result. CD4+ and CD8+T cells are required to obtain immunity against tumor.[65]

Reducing the Risk of Autoimmunity

Proper methods need to be followed for vaccination against cancer, to infuse a therapeutic antitumor response and avoid the undesirable height of autoimmune results. The peripheral immune system is occupied by a range of autoreactive T cells categorized into two different groups: low avidity T cells and low-to-high avidity T cells, tissue-specific factors avoided by central and peripheral tolerance. In high avidity, autoreactive T cells are prone to threat.^[66]

Evasion of Immune System

Tumor cells generally promote the activation of STAT1/ B7H1 and the secretion of IL-10 and TGF- β factors that hinder the antitumor response. The genetic changes like mutations occurring in the tumor antigens make the tumor cells less viable to immune recognition leading to immune rescue.^[66] The complication arises when mutations appear to begin in the antigen processing pathway like proteasome, TAP, and β 2- microglobulin.

Combination Therapy: Immunotherapy and Cancer Vaccines

The sensible expansion of the vaccines and the immunotherapeutic ways for the medication of cancer involves the tumor microenvironment and immune response that determine the antitumor immunity. The suppression of regulatory T cells (Treg) builds up a risk for the patients to establish autoimmune diseases (Fig. 4). The consolidation approach of vaccines and immunotherapy brings about the stimulation of inhibitory pathways in the immunosuppressive microenvironment of a tumor. A positive report on consolidation therapy has been found, which focuses on numerous arrays inside the immune system to increase immunity against tumors. The efficiency of the dendritic cell vaccine against B16 skin cancer in mice models can be increased by gene silencing of TGF- β 1, which decreases the regulatory T cells associated with tumor.^[67]



Figure 4. Future with the Combination use of cancer vaccines and immunotherapy to improve conditions against cancer.

The advantages of this consolidation approach overcome the immune checkpoint indicated in research studies that blockage of PD1/PD-L1 pathway by anti-PD-L1 to counteract the antibodies in addition to the exhaustion of regulatory T cells relapse the disease. The approach has proved one of the best therapies to suppress the tumor work against cancer.^[68]

The challenges faced in the development of cancer vaccines are tumor immune suppression and antigenicity. The obvious fact is the immune response of healthy individuals and cancer patients work differently. The cancer patients have to negotiate for both specific therapy and the tumor type. Antigenicity, where the vaccines do not have a specific target to tumor antigens, leads to mutations due to certain factors such as lifestyle, genetic, and environmental changes.^[69]

Nowadays, various new approaches are targeted toward immunotherapy and cancer vaccines, which include the combination of checkpoint inhibitors and personalized neoantigen vaccines. CTLA-4 inhibits the stimulation of T cells with the direct interaction of CD 80/86, where the T cell activation is stopped, leading to no immune response.^[70, 71] This means the blockage of checkpoints was done via the development of the monoclonal antibodies as an approach toward therapeutics. Ipilimumab, an FDAapproved monoclonal antibody against CTLA-4, is used for the treatment of melanoma.^[71, 72] In the previous research, it has been shown that checkpoint inhibitors have shown T cell responses and tumors carry huge mutational stress, which generates a lot of neoantigens.^[71] Melanoma and nonsmall lung cancer usually have a high load of neoantigens, which tend to show positive feedback against checkpoint inhibitors and a good overall survival rate. ^[72-74] On the contrary, tumors exhibiting fewer mutations such as thyroid cancer and leukemia show a low overall survival rate.^[75]

Another approach of cancer immunotherapy is to release the immune response through inhibition of checkpoint molecules with the use of inhibitors. Many of the patients do not respond well to immune checkpoint molecules, but they can benefit from the combination treatment of the inhibitors and antigen-specific therapy. CTLA-4, a checkpoint inhibitor, and Ipilimumab, the monoclonal antibody, have proved to lead NY-ESO-1 immune responses among patients with prostate, ovarian cancer, and melanoma.^[76, 77] There have been reports showing melanoma patients treated with NY-ESO-1 in combination with Nivolumab. PD-1 inhibitor showed a 25% positive response among the patients.^[78]

Future Perspectives of Combination Therapy

The most challenging task is to analyze precise dosage and efficient response in the combination of various check-point inhibitors including CTLA-4 and PD-1. Different components have been implemented for the blockage of PD-1/IDO/CTLA-4 pathways, which has shown encouraging results. This study combines the targeted therapies of the immune response with conventional therapies (Fig. 5) such as radiotherapy, chemotherapy, and chemically driven drugs to see the response on the cancer patients.^[79, 80]

There are many issues that raise a concern about the type of antigen, whether TAA or neoantigens. TAAs are most commonly classified by tumors, but the limiting factor is the tolerance of the immune response, whereas neoantigens are tedious, expensive, and it is difficult to know the tumor changes in a patient. There are clinical trials using a specific antigen for vaccine, but no trial has checked the combined effect of TAAs and neoantigens on the activation of the immune response. This needs future research.^[81]

Another aspect is combination therapy which involves the right therapies involved to have better results. It usually relies on the type of tumor, presence, and detection of biomarkers specific for patients. The use of vaccines applies as the last-line option. Therefore, to apply this process, we need to be sure about the dosage and the time for the immune response against a particular antigen.^[82]

Humans and animals have dissimilarities in their immune response. Genetically engineered mice, xenograft, and orthotopic models are available to avoid this complication. However, there is a demand for big animal models, but it raises a concern for their breeding, ethical rules, and housing.^[83]



Figure 5. Combination therapy for cancer treatment.

Conclusion

To find a cure for cancer, the future lies in the use of a combination approach, cancer vaccines and immunotherapy, in which the risk of side effects is being reduced as compared with other therapies such as chemotherapy, radiotherapy, and certain drugs available in the market. Efficiency as well as the survival rate of the patients suffering from cancer is increased. However, many studies need to be performed to find a cure for cancer. The combination of personalized therapies is making a new direction toward an individualized patient's immune response and microenvironment with new techniques for the treatment of cancer.

Disclosures

Ethics Committee Approval: Ethics committee approval was not requested for this study.

Peer-review: Externally peer-reviewed.

Conflict of Interest: None declared.

Authorship Contributions: Concept – T.G., M.G.; Design – W.P.S.; Supervision – W.P.S., U.G.; Materials – T.G., U.G.; Data collection &/or processing – T.G., M.M.; Analysis and/or interpretation – M.G., W.P.S.; Literature search – T.G., M.M.; Critical Review – U.G., M.G., N.K.P.

References

- 1. Vineis P, Wild CP. Global cancer patterns: causes and prevention. Lancet 2014;383:549–57.
- 2. Wild CP. The role of cancer research in noncommunicable disease control. J Natl Cancer Inst 2012;104:1051–8.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018;68:394–424.
- 4. Carioli G, Bertuccio P, Boffetta P, Levi F, La Vecchia C, Negri E, et al. European cancer mortality predictions for the year 2020 with a focus on prostate cancer. Ann Oncol 2020;31:650–8.
- 5. Hendrick RE, Baker JA, Helvie MA. Breast cancer deaths averted over 3 decades. Cancer 2019;125:1482–8.
- Arnold M, Sierra MS, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global patterns and trends in colorectal cancer incidence and mortality. Gut 2017;66:683–91.
- Lim RK, Kitts AB, Tremblay A. Lung cancer screening effective for reducing cancer deaths. Am Fam Physician 2020;101:70–1.
- Zheng R, Qu C, Zhang S, Zeng H, Sun K, Gu X, et al. Liver cancer incidence and mortality in China: Temporal trends and projections to 2030. Chin J Cancer Res 2018;30:571–9.
- Lortet-Tieulent J, Vaccarella S. International and subnational variation thyroid cancer incidence and mortality over 2008–2012. Revue d'Épidémiologie et de Santé Publique 2018;66:S254.
- 10. Rawla P, Sunkara T, Gaduputi V. Epidemiology of pancreatic

cancer: global trends, etiology and risk factors. World J Oncol 2019;10:10–27.

- Hoelzinger DB, Gendler SJ, Cohen PA, Dominguez AL, Smith SE, Lustgarten J, inventors; Mayo Foundation for Medical Education, assignee. Blocking IL-9 signaling in conjunction with chemotherapy to treat cancer. United States patent US 10,166,292. 2019 Jan 1. Available at: https://patents.google. com/patent/US9833512B2/en. Accessed Feb 4, 2022.
- 12. Goguet E, Klinman DM, Tross D. Intrapulmonary delivery of TLR agonists associated with systemic chemotherapy to treat metastatic cancer. J Immunol 2018;200.
- Inoue T, Katoh N, Ito YM, Kimura T, Nagata Y, Kuriyama K, et al. Stereotactic body radiotherapy to treat small lung lesions clinically diagnosed as primary lung cancer by radiological examination: A prospective observational study. Lung Cancer 2018;122:107–12.
- 14. Greenhalgh TA, Dearman C, Sharma RA. Combination of novel agents with radiotherapy to treat rectal cancer. Clin Oncol (R Coll Radiol) 2016;28:116–39.
- 15. Chen YW, Su YL, Hu SH, Chen SY. Functionalized graphene nanocomposites for enhancing photothermal therapy in tumor treatment. Adv Drug Deliv Rev 2016;105:190–204.
- Yamaguchi T, Imai M, Uematsu D. Hybrid approach using laparoscopy and transanal minimally invasive surgery to treat rectal cancer with invasion to the seminal vesicles. Asian J Endosc Surg 2017;10:219–22.
- 17. Butterfield LH. Cancer vaccines. BMJ 2015;350:h988.
- 18. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 2012;12:252–64.
- 19. Taube JM, Klein A, Brahmer JR, Xu H, Pan X, Kim JH, et al. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. Clin Cancer Res 2014;20:5064–74.
- 20. Salgado R, Denkert C, Demaria S, Sirtaine N, Klauschen F, Pruneri G, et al; International TILs Working Group 2014. The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: recommendations by an International TILs Working Group 2014. Ann Oncol 2015;26:259–71.
- 21. Zitvogel L, Kepp O, Kroemer G. Immune parameters affecting the efficacy of chemotherapeutic regimens. Nat Rev Clin Oncol 2011;8:151–60.
- 22. Pan RY, Chung WH, Chu MT, Chen SJ, Chen HC, Zheng L, et al. Recent development and clinical application of cancer vaccine: targeting neoantigens. J Immunol Res 2018;2018:4325874.
- Zhao X, Bose A, Komita H, Taylor JL, Chi N, Lowe DB, et al. Vaccines targeting tumor blood vessel antigens promote CD8(+) T cell-dependent tumor eradication or dormancy in HLA-A2 transgenic mice. J Immunol 2012;188:1782–8.
- 24. Kirkwood JM, Lee S, Moschos SJ, Albertini MR, Michalak JC, Sander C, et al. Immunogenicity and antitumor effects of vaccination with peptide vaccine+/-granulocyte-monocyte

colony-stimulating factor and/or IFN-alpha2b in advanced metastatic melanoma: Eastern Cooperative Oncology Group Phase II Trial E1696. Clin Cancer Res 2009;15:1443–51.

- Hailemichael Y, Dai Z, Jaffarzad N, Ye Y, Medina MA, Huang XF, et al. Persistent antigen at vaccination sites induces tumorspecific CD8+ T cell sequestration, dysfunction and deletion. Nat Med 2013;19:465–72.
- 26. Welters MJ, Kenter GG, Van Steenwijk PJ, Löwik MJ, Berendsvan Der Meer DM, Essahsah F, et al. Success or failure of vaccination for HPV16-positive vulvar lesions correlates with kinetics and phenotype of induced T-cell responses. PNAS 2010;107:11895–9.
- 27. Rosalia RA, Quakkelaar ED, Redeker A, Khan S, Camps M, Drijfhout JW, et al. Dendritic cells process synthetic long peptides better than whole protein, improving antigen presentation and T-cell activation. Eur J Immunol 2013;43:2554–65.
- 28. Palucka K, Banchereau J. Dendritic-cell-based therapeutic cancer vaccines. Immunity 2013;39:38–48.
- 29. Wierecky J, Müller MR, Wirths S, Halder-Oehler E, Dörfel D, Schmidt SM, et al. Immunologic and clinical responses after vaccinations with peptide-pulsed dendritic cells in metastatic renal cancer patients. Cancer Res 2006;66:5910–8.
- Musolino C, Allegra A, Innao V, Allegra AG, Pioggia G, Gangemi S. Inflammatory and anti-inflammatory equilibrium, proliferative and antiproliferative balance: the role of cytokines in multiple myeloma. Mediators Inflamm 2017;2017:1852517.
- O'Rourke MG, Johnson MK, Lanagan CM, See JL, O'Connor LE, Slater GJ, et al. Dendritic cell immunotherapy for stage IV melanoma. Melanoma Res 2007;17:316–22.
- 32. Kyte JA, Mu L, Aamdal S, Kvalheim G, Dueland S, Hauser M, et al. Phase I/II trial of melanoma therapy with dendritic cells transfected with autologous tumor-mRNA. Cancer Gene Ther 2006;13:905–18.
- 33. Berntsen A, Trepiakas R, Wenandy L, Geertsen PF, thor Straten P, Andersen MH, et al. Therapeutic dendritic cell vaccination of patients with metastatic renal cell carcinoma: a clinical phase 1/2 trial. J Immunother 2008;31:771–80.
- 34. Avigan DE, Vasir B, George DJ, Oh WK, Atkins MB, McDermott DF, et al. Phase I/II study of vaccination with electrofused allogeneic dendritic cells/autologous tumor-derived cells in patients with stage IV renal cell carcinoma. J Immunother 2007;30:749–61.
- 35. Babatz J, Röllig C, Löbel B, Folprecht G, Haack M, Günther H, et al. Induction of cellular immune responses against carcinoembryonic antigen in patients with metastatic tumors after vaccination with altered peptide ligand-loaded dendritic cells. Cancer Immunol Immunother 2006;55:268–76.
- 36. Loveland BE, Zhao A, White S, Gan H, Hamilton K, Xing PX, et al. Mannan-MUC1-pulsed dendritic cell immunotherapy: a phase I trial in patients with adenocarcinoma. Clin Cancer Res 2006;12:869–77.

- 37. Morse MA, Clay TM, Hobeika AC, Osada T, Khan S, Chui S, et al. Phase I study of immunization with dendritic cells modified with fowlpox encoding carcinoembryonic antigen and costimulatory molecules. Clin Cancer Res 2005;11:3017–24.
- 38. Soiffer R, Hodi FS, Haluska F, Jung K, Gillessen S, Singer S, et al. Vaccination with irradiated, autologous melanoma cells engineered to secrete granulocyte-macrophage colony-stimulating factor by adenoviral-mediated gene transfer augments antitumor immunity in patients with metastatic melanoma. J Clin Oncol 2003;21:3343–50.
- 39. Luiten RM, Kueter EW, Mooi W, Gallee MP, Rankin EM, Gerritsen WR, et al. Immunogenicity, including vitiligo, and feasibility of vaccination with autologous GM-CSF-transduced tumor cells in metastatic melanoma patients. J Clin Oncol 2005;23:8978– 91.
- 40. Le DT, Brockstedt DG, Nir-Paz R, Hampl J, Mathur S, Nemunaitis J, et al. A live-attenuated Listeria vaccine (ANZ-100) and a live-attenuated Listeria vaccine expressing mesothelin (CRS-207) for advanced cancers: phase I studies of safety and immune induction. Clin Cancer Res 2012;18:858–68.
- 41. Barth RJ Jr, Fisher DA, Wallace PK, Channon JY, Noelle RJ, Gui J, et al. A randomized trial of ex vivo CD40L activation of a dendritic cell vaccine in colorectal cancer patients: tumor-specific immune responses are associated with improved survival. Clin Cancer Res 2010;16:5548–56.
- 42. Harrop R, Connolly N, Redchenko I, Valle J, Saunders M, Ryan MG, et al. Vaccination of colorectal cancer patients with modified vaccinia Ankara delivering the tumor antigen 5T4 (Tro-Vax) induces immune responses which correlate with disease control: a phase I/II trial. Clin Cancer Res 2006;12:3416–24.
- 43. Kim TS, Chopra A, O-Sullivan IS, Cohen EP. Enhanced immunity to breast cancer in mice immunized with fibroblasts transfected with a complementary DNA expression library from breast cancer cells: Enrichment of the vaccine for immunotherapeutic cells. J Immunother 2006;29:261–73.
- 44. Schumacher L, Ribas A, Dissette VB, McBride WH, Mukherji B, Economou JS, et al. Human dendritic cell maturation by adenovirus transduction enhances tumor antigen-specific T-cell responses. J Immunother 2004;27:191–200.
- 45. Kaufman HL, Bines SD. OPTIM trial: a Phase III trial of an oncolytic herpes virus encoding GM-CSF for unresectable stage III or IV melanoma. Future Oncol 2010;6:941–9.
- 46. Lindsey KR, Gritz L, Sherry R, Abati A, Fetsch PA, Goldfeder LC, et al. Evaluation of prime/boost regimens using recombinant poxvirus/tyrosinase vaccines for the treatment of patients with metastatic melanoma. Clin Cancer Res 2006;12:2526–37.
- 47. Amato RJ, Shingler W, Goonewardena M, de Belin J, Naylor S, Jac J, et al. Vaccination of renal cell cancer patients with modified vaccinia Ankara delivering the tumor antigen 5T4 (TroVax) alone or administered in combination with interferon-alpha (IFN-alpha): a phase 2 trial. J Immunother 2009;32:765–72.

- 48. Arlen PM, Skarupa L, Pazdur M, Seetharam M, Tsang KY, Grosenbach DW, et al. Clinical safety of a viral vector based prostate cancer vaccine strategy. J Urol 2007;178:1515–20.
- 49. Marshall JL, Gulley JL, Arlen PM, Beetham PK, Tsang KY, Slack R, et al. Phase I study of sequential vaccinations with fowlpox-CEA(6D)-TRICOM alone and sequentially with vaccinia-CEA(6D)-TRICOM, with and without granulocyte-macrophage colony-stimulating factor, in patients with carcinoembryonic antigen-expressing carcinomas. J Clin Oncol 2005;23:720–31.
- Nicholaou T, Ebert LM, Davis ID, McArthur GA, Jackson H, Dimopoulos N, et al. Regulatory T-cell-mediated attenuation of T-cell responses to the NY-ESO-1 ISCOMATRIX vaccine in patients with advanced malignant melanoma. Clin Cancer Res 2009;15:2166–73.
- 51. Motohashi S, Nagato K, Kunii N, Yamamoto H, Yamasaki K, Okita K, et al. A phase I-II study of alpha-galactosylceramidepulsed IL-2/GM-CSF-cultured peripheral blood mononuclear cells in patients with advanced and recurrent non-small cell lung cancer. J Immunol 2009;182:2492–501.
- 52. Nemunaitis J, Nemunaitis M, Senzer N, Snitz P, Bedell C, Kumar P, et al. Phase II trial of Belagenpumatucel-L, a TGF-beta2 antisense gene modified allogeneic tumor vaccine in advanced non small cell lung cancer (NSCLC) patients. Cancer Gene Ther 2009;16:620–4.
- 53. Nemunaitis J, Jahan T, Ross H, Sterman D, Richards D, Fox B, et al. Phase 1/2 trial of autologous tumor mixed with an allogeneic GVAX vaccine in advanced-stage non-small-cell lung cancer. Cancer Gene Ther 2006;13:555–62.
- 54. Okaji Y, Tsuno NH, Tanaka M, Yoneyama S, Matsuhashi M, Kitayama J, et al. Pilot study of anti-angiogenic vaccine using fixed whole endothelium in patients with progressive malignancy after failure of conventional therapy. Eur J Cancer 2008;44:383–90.
- 55. Powell A, Creaney J, Broomfield S, Van Bruggen I, Robinson B. Recombinant GM-CSF plus autologous tumor cells as a vaccine for patients with mesothelioma. Lung Cancer 2006;52:189–97.
- 56. Fakhrai H, Mantil JC, Liu L, Nicholson GL, Murphy-Satter CS, Ruppert J, et al. Phase I clinical trial of a TGF-beta antisensemodified tumor cell vaccine in patients with advanced glioma. Cancer Gene Ther 2006;13:1052–60.
- 57. Victora GD, Socorro-Silva A, Volsi EC, Abdallah K, Lima FD, Smith RB, et al. Immune response to vaccination with DNA-Hsp65 in a phase I clinical trial with head and neck cancer patients. Cancer Gene Ther 2009;16:598–608.
- Zerbini A, Pilli M, Laccabue D, Pelosi G, Molinari A, Negri E, et al. Radiofrequency thermal ablation for hepatocellular carcinoma stimulates autologous NK-cell response. Gastroenterology 2010;138:1931–42.
- 59. Gravante G, Sconocchia G, Ong SL, Dennison AR, Lloyd DM. Immunoregulatory effects of liver ablation therapies for the

treatment of primary and metastatic liver malignancies. Liver Int 2009;29:18–24.

- 60. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al; IMPACT Study Investigators. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. N Engl J Med 2010;363:411–22.
- 61. Gilboa E. The promise of cancer vaccines. Nat Rev Cancer 2004;4:401–11.
- 62. Zeis M, Siegel S, Wagner A, Schmitz M, Marget M, Kühl-Burmeister R, et al. Generation of cytotoxic responses in mice and human individuals against hematological malignancies using survivin-RNA-transfected dendritic cells. J Immunol 2003;170:5391–7.
- 63. Andersen MH, Pedersen LO, Capeller B, Bröcker EB, Becker JC, thor Straten P. Spontaneous cytotoxic T-cell responses against survivin-derived MHC class I-restricted T-cell epitopes in situ as well as ex vivo in cancer patients. Cancer Res 2001;61:5964– 8.
- 64. Pardoll D. Does the immune system see tumors as foreign or self? Annu Rev Immunol 2003;21:807–39.
- 65. Kwon B, Lee HW, Kwon BS. New insights into the role of 4-1BB in immune responses: beyond CD8+ T cells. Trends Immunol 2002;23:378–80.
- 66. Yee C, Thompson JA, Byrd D, Riddell SR, Roche P, Celis E, et al. Adoptive T cell therapy using antigen-specific CD8+ T cell clones for the treatment of patients with metastatic melanoma: in vivo persistence, migration, and antitumor effect of transferred T cells. Proc Natl Acad Sci U S A 2002;99:16168–73.
- 67. Conroy H, Galvin KC, Higgins SC, Mills KH. Gene silencing of TGF-β1 enhances antitumor immunity induced with a dendritic cell vaccine by reducing tumor-associated regulatory T cells. Cancer Immunol Immunother 2012;61:425–31.
- Goding SR, Wilson KA, Xie Y, Harris KM, Baxi A, Akpinarli A, et al. Restoring immune function of tumor-specific CD4+ T cells during recurrence of melanoma. J Immunol 2013;190:4899– 909.
- 69. Bowen WS, Svrivastava AK, Batra L, Barsoumian H, Shirwan H. Current challenges for cancer vaccine adjuvant development. Expert Rev Vaccines 2018;17:207–15.
- 70. Aldous AR, Dong JZ. Personalized neoantigen vaccines: A new approach to cancer immunotherapy. Bioorg Med Chem 2018;26:2842–9.
- 71. Ward JP, Gubin MM, Schreiber RD. The role of neoantigens in naturally occurring and therapeutically induced immune responses to cancer. Adv Immunol 2016;130:25–74.
- 72. Katsnelson A. Mutations as munitions: Neoantigen vaccines get a closer look as cancer treatment. Nat Med 2016;22:122–4.
- 73. Schumacher TN, Hacohen N. Neoantigens encoded in the cancer genome. Curr Opin Immunol 2016;41:98–103.
- 74. Shukla SA, Howitt BE, Wu CJ, Konstantinopoulos PA. Predicted neoantigen load in non-hypermutated endometrial cancers:

Correlation with outcome and tumor-specific genomic alterations. Gynecol Oncol Rep 2016;19:42–5.

- 75. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, et al; Australian Pancreatic Cancer Genome Initiative. Signatures of mutational processes in human cancer. Nature 2013;500:415–21.
- 76. Sioud M, Nyakas M, Sæbøe-Larssen S, Mobergslien A, Aamdal S, Kvalheim G. Diversification of antitumour immunity in a patient with metastatic melanoma treated with ipilimumab and an IDO-silenced dendritic cell vaccine. Case Rep Med 2016;2016:9639585.
- 77. Dos Santos LI, Galvao-Filho B, de Faria PC, Junqueira C, Dutra MS, Teixeira SM, et al. Blockade of CTLA-4 promotes the development of effector CD8+ T lymphocytes and the therapeutic effect of vaccination with an attenuated protozoan expressing NY-ESO-1. Cancer Immunol Immunother 2015;64:311–23.
- 78. Weber JS, Kudchadkar RR, Gibney GT, De Conti RC, Yu B, Wang W, et al. Phase I/II trial of PD-1 antibody nivolumab with peptide vaccine in patients naive to or that failed ipilimumab. ASCO Meeting Abstracts 2013;31:9011.

- 79. Ishihara D, Pop L, Takeshima T, Iyengar P, Hannan R. Rationale and evidence to combine radiation therapy and immunotherapy for cancer treatment. Cancer Immunol Immunother 2017;66:281–98.
- 80. Cook AM, Lesterhuis WJ, Nowak AK, Lake RA. Chemotherapy and immunotherapy: mapping the road ahead. Curr Opin Immunol 2016;39:23–9.
- Aurisicchio L, Salvatori E, Lione L, Bandini S, Pallocca M, Maggio R, et al. Poly-specific neoantigen-targeted cancer vaccines delay patient derived tumor growth. J Exp Clin Cancer Res 2019;38:78.
- Sahin U, Derhovanessian E, Miller M, Kloke BP, Simon P, Löwer M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. Nature 2017;547:222–6.
- 83. Overgaard NH, Fan TM, Schachtschneider KM, Principe DR, Schook LB, Jungersen G. Of mice, dogs, pigs, and men: choosing the appropriate model for immuno-oncology research. ILAR J 2018;59:247–62.